

SPlink HRP Mouse Detection (DAB) Kit

(Horseradish peroxidase labeled-streptavidin-biotin detection system for mouse antibody with DAB chromogen)

| Storage: 2-8°C | Catalog No.: | D02-110 D02-18 | 110ml 🗌 18 ml 🗍 |
|----------------|--------------|-------------------|--------------------|
| Storage. 2-0 C | | D02-10 | 6 ml |

Intended Use:

SPlink HRP Mouse Detection (DAB) kit is intended for using with mouse primary antibody (user-supplied) to detect the presence of antigens in human tissue or cell preparations under light microscopy. Most commonly used specimens for this system are: frozen tissue, paraffin-embedded tissue, freshly prepared lymphocytes and fixed culture cells.

Horseradish peroxidase (HRP) labeled-streptavidin and biotinylated secondary antibody amplification system has become a standard technique in immunochemical staining^{1,2}. SPlink HRP Mouse Detection (DAB) kit uses human-absorbed, biotinylated, affinity-purified secondary antibody reacts with the user supplied primary antibody bound to the specific epitope of the antigen in tissue or cell. Horseradish peroxidase (HRP) labeled streptavidin then reacts with biotinylated secondary antibody to form a HRP-streptavidin-biotin complex. The HRP enzyme of the streptavidin complex catalyzed the substrate/chomogen, 3,3qdiaminobenzidine (DAB substrate) reaction to form brown color deposit at the antigen site. The antigen then can be visualized under microscope. Compared to traditional ABC method which uses avidin, SPlink HRP Mouse Detection (DAB) kit demonstrates stronger binding strength to bind biotin and less non-specific background staining. Pre-Blocking Solution in the kit will help to eliminate non-specific background.

Higher sensitivity and lower background give SPlink HRP Mouse Detection (DAB) kit a higher signal-noise ratio. More than sufficient volume of DAB chormogen is provided in the kit so that customers may use 2 drops of DAB chromogen per ml to obtain higher sensitivity and contrast.

Kit Components:

| No. | | 1 | 2 | 3 | 4A | 4B |
|------------|---------------------------------|--------------------------|------------------------------------------------|------------------------------------------|---------------------------------|------------------------------------|
| Catalog No | Name | Pre-Blocking Solution | Biotinylated anti- mouse second antibody | Streptavidin- peroxidase conjugate | DAB substrate (Ready-to-use) | DAB chromogen (Concentrated) |
| D02-6 | SPlink HRP Mouse DAB 6ml kit | 6ml | 6 ml | 6 ml | 12 ml | 1.5ml |
| D02-18 | SPlink HRP Mouse DAB 6ml kit | 18 ml | 18 ml | 18 ml | 15ml x 2 | 2 ml |
| D02-110 | SPlink HRP Mouse Bulk kit | 110ml | 110ml | 110ml | Not included | Not included |

Recommended Protocol:

- 1. Fixation: To ensure the quality of the staining and obtain reproducible performance, user needs to supply appropriately fixed tissue and well prepared slides.
- 2. Tissue need to be adhered to the slide tightly to avoid tissue falling off.
- 3. Paraffin embedded section must be deparffinized with xylene and rehydrated with a graded series of ethanol before staining.
- 4. Cell smear samples should be made as much monolayer as possible to obtain satisfactory results.
- 5. Three control slides will aid the interpretation of the result: positive tissue control, reagent control (slide treated with Isotype control reagent), and negative control.
- 6. Start staining procedures: DO NOT let specimen or tissue dry from this point on.

| Reagent | Staining Procedures | Incubation Time (Min.) |
|---------------------------------------|----------------------------------------------------------------------------------------|---------------------------|
| Peroxidase blocking reagent: | a. Apply 2 drops (100 L) or enough volume of Peroxidase | 10 min. |
| Supplied by user. | blocking reagent (Ready-to-use 3% H ₂ O ₂ solution) to cover the | |
| | tissue section and incubate | |
| | b. Rinse the slide using distilled water. | |
| 2. HIER Pretreatment: refer to | a. Heat Induced Epitope Retrieval (HIER) may be required for | |
| antibody spec. sheet | primary antibody suggested by vendor | |
| | b. Wash with PBS 2 min., 3 times. | |
| 3. Reagent 1: | a. Add 2 drops or enough volume of Pre-blocking Solution to | 10 min. |
| Pre-blocking Solution | completely cover the tissue section and Incubate | |
| - | b. Blot off solution. DO NOT RINSE. | |
| 4. Primary antibody: | a. Apply 2 drops or enough volume of Primary antibody to cover | 30-60 min. |
| Supplied by user. Investigator | the tissue section completely. Incubate in moist chamber for 30- | |
| needs to optimize dilution and | 60 min. | |
| incubation time. | b. Rinse with PBS for 2 min., 3 times. | |
| 5. Reagent 2: | a. Apply 2 drops or enough volume of secondary antibody to | 10 min. |
| Ready to use Secondary antibody | cover the tissue section completely and incubate. | |
| | b. Rinse with PBS for 2 min., 3 times. | |
| 6. Reagent 3: | a. Apply 2 drops or enough volume of HRP-Streptavidin to cover | 10 min. |
| Ready to use HRP-Streptavidin | the tissue section completely and incubate. | |
| | b. Rinse with PBS for 2 min., 3 times. | |
| 7. Reagents 4A, 4B: | a. Add 1 drop or 2 drops (for higher sensitivity and contrast) of | 5 min. |
| 4A: DAB Substrate (RTU) | Reagent 4B to 1ml of 4A. Mix well. Protect from light and use | |
| 4B : DAB Chromogen Concentrate | within 5 hours. | |
| | b. Apply 2 drops (100 L) or enough volume of pre-mixed DAB | |
| | chromogen to completely cover tissue and Incubate 5 minutes. | |
| | c. Rinse with distill water for 2 min, 3 times. | |
| 8. Hematoxylin: | a. Counterstain with 2 drops (100 ul) or more drops to cover | |
| Supplied by user | tissue completely and wait about 10-20 seconds. | |
| | b. Rinse thoroughly under tap water for 1-2 min. | |
| | c. Put slides in PBS until show blue color (about 30-60 seconds) | |
| | d. Rinse well in distiled water | |
| 9. Mounting media: | Follow the manufacture data sheet procedure for mounting. | |
| Supplied by user | Recommended product: | |
| | O-Mount: Cat. No. E02-15 (15ml) | |
| | Simpo-Mount: Cat.No. E03-15 (15ml) or E03-100 (100ml) | |

Protocol Notes:

- 1. The fixation, tissue slide thickness, antigen retrieval and primary antibody dilution and incubation time effect results significantly. Investigator needs to consider all factors and determine optimal conditions when interpret the result.
- 2. Tissue staining is dependent upon the proper handling and processing of tissues prior to staining. Improper tissue preparation may lead to false negative results or inconsistent results.
- 3. Do not mix reagents from different lot.
- 4. Do not allow the slides to dry at any time during staining

Related Products:

| Product | Catalog No. | Size | Product | Catalog No. | Size |
|----------------------------|----------------|------------|--------------------------------|-------------|--------|
| SPlink HRP Broad Bulk kit | D01-110 | 110ml | Simplified Streptavidin HRP | D30-1 | 1ml |
| | | | Rabbit concentrate kit (1:100) | | |
| SPlink HRP Broad DAB Kit | D01-18 / D01-6 | 18ml / 6ml | Simplified Streptavidin HRP | D31-1 | 1ml |
| | | | Mouse concentrate kit (1:100) | | |
| SPlink HRP Rabbit Bulk kit | D03-110 | 110ml | Streptavidin Peroxidase (RTU) | D25-110/ | 110ml |
| | | | | D25-18 | 18ml |
| SPlink HRP Rabbit DAB Kit | D03-18 / D03-6 | 18ml / 6ml | SPlink HRP Broad AEC | D04-18 / | 18ml / |
| | | | | D04-6 | 6ml |
| SPlink HRP Goat Bulk kit | D76-110 | 110ml | SPlink HRP Mouse AEC | D05-18 / | 18ml / |
| | | | | D05-6 | 6ml |
| SPlink HRP Goat DAB Kit | D76-18 / D76-6 | 18ml / 6ml | SPlink HRP Rabbit AEC | D06-18 / | 18ml / |
| | | | | D06-6 | 6ml |

Precautious:

DAB may be carcinogenic. Handle all specimens as potential infectious materials, wear gloves and protection cloth.

Remarks:

For research use or investigation only. Not for diagnostic or therapeutic use.

References:

- 1. Elias, J.M. et al. Sensitivity and Detection Efficiency of the Peroxidase antiperoxidase (PAP) Avidin-Biotin Peroxidase Complex (ABC), and Peroxidase-Labeled Avidin-Biotin (LAB Methods. AM J Clin Pathol 92:62-67, 1989.
- 2. Polak, J.M and Van Noorden, S. Introduction to Immunocytochemistry Second Edition. Bios Scientific Publishers. 41-54. 1997.